

DYSOXYLIN, A LIMONOID FROM *DYSOXYLUM RICHII*

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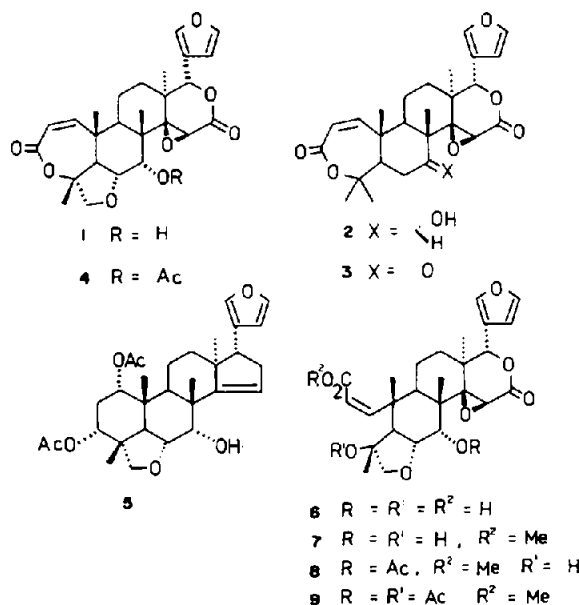
Abstract—Dysoxylin, a new limonoid, was isolated from the fresh leaves of *Dysoxylum richii* collected in Suva, Fiji. The proposed structure was based on spectral assignments and chemical interconversions.

INTRODUCTION

Limonoids are widely distributed in plants of the Meliaceae [1]. We have examined methanolic extracts of the leaves of *Dysoxylum richii* (Gray) C.D.C. as part of a study of the chemical constituents of Fijian plants, and we now wish to report the structure of dysoxylin (1), the second limonoid from a plant in the genus *Dysoxylum* [2].

RESULTS AND DISCUSSION

The chloroform-soluble portion of the leaf extract was fractionated by silica gel flash chromatography to give dysoxylin (1) as an amorphous solid (yield 0.1% dry wt)



that had a molecular formula of C₂₆H₃₀O₈ (M⁺ 470.1937 dev. – 0.4 mmu). Bands at 3480, 1740, and 1690 cm^{–1} in the IR spectrum of 1 could be assigned to hydroxyl, saturated lactone and unsaturated lactone functionalities respectively.

The ¹H (Table 1) and ¹³C (Table 2) NMR spectra of dysoxylin (1) indicated that it was a limonoid and that it contained A, C, and D ring functionality identical to that found in obacunol (2) [3] and obacunone (3) [4]. Resonances at δ3.01 (d, J = 12 Hz, 1H), 4.01 (dd, J = 12, 2 Hz, 1H) and 3.64 (d, J = 2 Hz, 1H) in the ¹H NMR could be assigned to methine protons on C5, C6, and C7 of the B ring of the limonoid skeleton. Acetylation of dysoxylin (Ac₂O/DMAP/triethylamine [5]) gave a good yield of the monoacetate (4). The ¹H NMR of 4 showed that H-7 had undergone an acetylation shift from δ3.64 in 1 to 5.0 in 4, and that H15 had shifted upfield from δ3.95 in 1 to 3.69 in 4. These data were consistent with the presence of an alpha hydroxyl substituent at C7 in dysoxylin.

Only four tertiary methyl resonances were apparent in the ¹H and ¹³C NMR spectra of 1 (Tables 1 and 2), in contrast to the five expected for the limonoid skeleton and observed in obacunol (2) and obacunone (3). A pair of geminal proton resonances at δ4.33 (d, J = 9 Hz) and 3.82 (d, J = 9 Hz) indicated that the fifth methyl residue was connected to C6 via an ether linkage identical to that found in salannin [6] and diacetylvilasinin (5) [7].

The stereochemistry of dysoxylin (1) was determined by an analysis of spin–spin coupling constants and a series of NOE experiments. A 12 Hz vicinal coupling constant between H5 and H6 suggested that ring B was in a chair conformation and that both protons were axial, while a 2 Hz coupling between H6 and H7 implied that H7 was equatorial. Irradiation of the C19 methyl resonance in the monoacetate 4 induced NOE's into Me-30, Me-29 and H6. Irradiation of the C29 methyl resonance induced NOE's into Me-19, H-6, and H-28, β, while irradiation of the C18 methyl resonance induced an NOE into H9. These results are consistent with the stereochemistry shown for 1.

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Table 1. ^1H NMR data for dysoxylin (1), obacunol (2), dysoxylin acetate (4) and 1,3-diacetylvilasinin (5) (The spectra of 1 and 4 were run in CDCl_3 at 400 MHz, chemical shifts are reported in ppm from internal TMS)

Compound	1	2*	4	5†
Protons on carbon no.				
1	6.33 <i>d</i> 12 Hz	6.57 <i>d</i> 12 Hz	6.31 <i>d</i> 12 Hz	
2	5.85 <i>d</i> 12 Hz	5.89 <i>d</i> 12 Hz	5.92 <i>d</i> 12 Hz	
5	3.01 <i>d</i> 12 Hz		2.82 <i>d</i> 12 Hz	2.67 <i>d</i> 12.5 Hz
6	4.01 <i>dd</i> 12 and 2 Hz		4.07 <i>dd</i> 12 and 2 Hz	4.14 <i>dd</i> 12.5 and 3 Hz
7	3.64 <i>d</i> 2 Hz	3.52 <i>t</i> 2 Hz	5.01 <i>d</i> 2 Hz	4.18 <i>d</i> 3 Hz
9	2.48 <i>t</i> 8 Hz		2.47 <i>t</i> 8 Hz	2.56 <i>m</i>
15	3.95 <i>s</i>	3.90 <i>s</i>	3.69 <i>s</i>	
17	5.58 <i>s</i>	5.64 <i>s</i>	5.58 <i>s</i>	
18	1.21 <i>s</i>	1.25 <i>s</i> ‡	1.20 <i>s</i>	0.84 <i>s</i>
19	1.28 <i>s</i>	1.30 <i>s</i> ‡	1.29 <i>s</i>	0.97 <i>s</i>
21	7.39 <i>m</i>	7.49 <i>d</i> 1 Hz	7.41 <i>m</i>	7.36 <i>m</i>
22	6.33 <i>d</i> 1 Hz	6.42 <i>t</i> 1 Hz	6.34 <i>d</i> 1 Hz	6.27 <i>m</i>
23	7.39 <i>m</i>	7.49 <i>d</i> 1 Hz	7.41 <i>m</i>	7.24 <i>m</i>
28	3.82 <i>d</i> 9 Hz	1.25 <i>s</i>	3.77 <i>d</i> 9 Hz	3.59 <i>d</i> 7 Hz
	4.33 <i>d</i> 9 Hz		4.24 <i>d</i> 9 Hz	3.56 <i>d</i> 7 Hz
29	1.60 <i>s</i>	1.43 <i>s</i> ‡	1.55 <i>s</i>	1.18 <i>s</i>
30	1.08 <i>s</i>	1.05 <i>s</i> ‡	1.17 <i>s</i>	1.11 <i>s</i>

* Data from ref. [3].

† See ref. [7] for complete data.

‡ Unassigned.

Dysoxylin (1) was hydrolysed with 0.05M NaOH in dioxane to give the dihydroxy acid 6, which was methylated with diazomethane to give the ester 7. Acetylation of ester 7 (Ac_2O /pyridine) gave a mixture of mono- and diacetates 8 and 9. The spectral data observed for all the transformation products were consistent with the proposed structures.

EXPERIMENTAL

Isolation and extraction. Fresh leaves of *Dysoxylum richii* were collected from Suva, Fiji in December 1985. A voucher specimen of this plant is deposited at the South Pacific Regional Herbarium, Institute of Natural Resources, University of the South Pacific, Suva, Fiji. The air-dried leaves (300 g) were soaked in MeOH (1.5 l) for 5 days. The MeOH extract was concd to a gum, H_2O was added and the resulting suspension was extracted successively with petrol (bp 60–80°) and CHCl_3 . The concd CHCl_3 extract was dissolved in EtOAc (10 ml), soaked over alumina (10 g), filtered, and concd to give a gum (2.4 g). Fractionation of the gum by flash chromatography on silica gel using mixtures of cyclohexane–EtOAc gave crude dysoxylin (1) (0.37 g). Preparative TLC (1:1 toluene/EtOAc) of the crude material gave 1 as solid, mp 247–251°, $[\alpha]_D^{25} + 72.5^\circ$ (CHCl_3 ; c 0.15); $\text{C}_{26}\text{H}_{30}\text{O}_8$ EIMS $[M]^+$ 470.1937 (dev. -0.4 mmu); ^1H NMR see Table 1; ^{13}C NMR see Table 2.

Dysoxylin acetate (4). To a stirred mixture of 1 (20 mg) and NEt_3 (0.5 ml) was added 4-(*N,N*-dimethylamino)pyridine (DMAP) (3 mg). The reaction mixture was heated to 60° for 3 hr.

After cooling, the reaction mixture was diluted with water (5 ml), then extracted with CHCl_3 . The organic layer was washed sequentially with aq. HCl (5%) and aq. NaHCO_3 (5%) before drying and concentration *in vacuo* to give dysoxylin acetate (4) as a solid, mp 235–245°; $\text{C}_{28}\text{H}_{32}\text{O}_9$ EIMS M^+ 512.2050 (dev. $+0.3$ mmu); ^1H NMR see Table 1; ^{13}C NMR see Table 2.

Dihydroxy acid. To a solution of 1 (3 mg) in dioxane (0.5 ml) was added 0.05 M NaOH (0.5 ml). The solution was refluxed for 30 min, then warmed at 50° for 90 min. Work-up gave the dihydroxyacid 6. ^1H NMR (CDCl_3) 1.05 (3H, *s*, Me-30), 1.27 (3H, *s*, Me-18), 1.34 (3H, *s*, Me-19), 1.45 (3H, *s*, Me-29).

Methyl ester. Dihydroxyacid 6 was methylated with diazomethane to give the methyl ester 7. $\text{C}_{27}\text{H}_{34}\text{O}_9$ EIMS $[M]^+$ 502.2110 (dev. 0.7 mmu); ^1H NMR (CDCl_3) 1.07 (3H, *s*, Me-30), 1.22 (3H, *s*, Me-18), 1.35 (3H, *s*, Me-19), 1.45 (3H, *s*, Me-29), 3.75 (3H, *s*, COOMe).

Monoacetate and diacetate. Acetylation of 7 using Ac_2O –pyr. (room temp. for 24 hr) gave the monoacetate 8 and the diacetate 9 as gums. 8: ^1H NMR (CDCl_3) 1.11, 1.21, 1.32 and 1.45 (3H each, *s*, Me-30, Me-18, Me-19 and Me-29) 2.20 (3H, *s*, COMe), 3.73 (3H, *s*, COOMe), 5.00 ppm (1H, *d*, $J = 2$ Hz, H-7); 9: ^1H NMR (CDCl_3) 1.12, 1.23, 1.33 and 1.5 (+ H_2O) (3H each, *s*, Me-30, Me-18, Me-19 and Me-29), 1.93 and 2.20 (3H each, *s*, COMe), 3.70 ppm (3H, *s*, COOMe).

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Table 2. ^{13}C NMR data. All spectra were run in CDCl_3 (Chemical shifts are in ppm from internal TMS)

Carbon	1*	3†	4	5‡
1	149.8	156.8	149.3	72.28
2	119.4	122.7	119.9	27.66
3	167.7§	166.7	166.8§	71.76
4	84.9	84.0	84.4	42.32
5	52.4	53.3¶	54.3	39.62
6	77.5	39.0	76.0	72.90
7	69.6	207.5	71.0	74.01
8	43.3	52.9	43.3	45.80
9	40.4	49.1	42.0	33.62
10	44.8	43.1	44.5	39.20
11	15.4	17.0	15.4	15.21
12	26.0	32.6	26.0	33.02
13	38.5	37.3	38.8	47.36
14	69.6	65.1	69.5	159.88
15	58.1	57.2¶	56.1	120.75
16	165.6§	166.9	165.5§	34.35
17	78.1	77.9	78.0	51.58
18	17.5	16.4	17.7§	21.16
19	17.2	19.4	17.3§	26.20
20	120.5	120.0	120.2	124.52
21	143.0	143.1	143.1	139.76
22	109.9	109.7	109.8	111.06
23	141.2	140.9	141.2	142.62
28	81.3	21.0	81.3	77.88
29	23.5	26.7	23.4§	15.37
30	17.9	32.0	18.5§	19.47
MeCO	—	—	21.0	21.16, 21.09
MeCO	—	—	170.0	169.97, 170.28

* Assignments were based on comparison to literature values for model compounds, APT and one bond HETCOR.

† Data from ref. [4] (see also [8]).

‡ Data from ref. [7].

§ Values in same vertical column may be reversed.

|| Unassigned.

¶ Assignments are reversed in ref. [4].

REFERENCES

1. Taylor, D. A. H. (1984) in *Progress in the Chemistry of Organic Natural Products* (Herz, W., Grisebach, H. and Kirby, G. W., eds) Vol. 45, p. 1. Springer, New York.
2. Singh, S., Garg, H. S. and Khanna, N. M. (1976) *Phytochemistry* 15, 2001.
3. Dreyer, D. L. (1968) *J. Org. Chem.* 33, 3577.
4. Dreyer, D. L. and Bennett, R. D. (1976) *Tetrahedron* 32, 2367.
5. Hasner, A., Krepski, L. R. and Alexanian, V. (1978) *Tetrahedron* 34, 2069.
6. Henderson, R., McCrindle, R., Melera, A. and Overton, K. H. (1968) *Tetrahedron* 24, 1525.
7. Kraus, W. and Cramer, R. (1981) *Liebigs. Ann. Chem.* 181.
8. Kubo, I., Tanis, S. P., Lee, Y.-W., Miura, I., Nakanishi, K. and Chapya, A. (1976) *Heterocycles* 5, 485.