

## DYSOXYLIN, A LIMONOID FROM *DYSOXYLUM RICHII*

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**Key Word Index**—*Dysoxylum richii*; Meliaceae; limonoid; dysoxylin; tetranortriterpene.

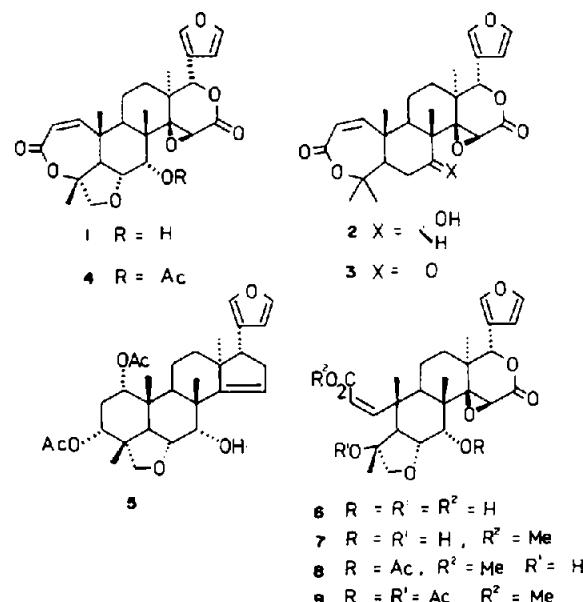
**Abstract**—Dysoxylin, a new limonoid, was isolated from the fresh leaves of *Dysoxylum richii* collected in Suva, Fiji. The proposed structure was based on spectral assignments and chemical interconversions.

### INTRODUCTION

Limonoids are widely distributed in plants of the Meliaceae [1]. We have examined methanolic extracts of the leaves of *Dysoxylum richii* (Gray) C.DC. as part of a study of the chemical constituents of Fijian plants, and we now wish to report the structure of dysoxylin (1), the second limonoid from a plant in the genus *Dysoxylum* [2].

### RESULTS AND DISCUSSION

The chloroform-soluble portion of the leaf extract was fractionated by silica gel flash chromatography to give dysoxylin (1) as an amorphous solid (yield 0.1% dry wt).



that had a molecular formula of  $C_{26}H_{30}O_8$  ( $M^+$  470.1937 dev. - 0.4 mmu). Bands at 3480, 1740, and  $1690\text{ cm}^{-1}$  in the IR spectrum of 1 could be assigned to hydroxyl, saturated lactone and unsaturated lactone functionalities respectively.

The  $^1\text{H}$  (Table 1) and  $^{13}\text{C}$  (Table 2) NMR spectra of dysoxylin (1) indicated that it was a limonoid and that it contained A, C, and D ring functionality identical to that found in obacunol (2) [3] and obacunone (3) [4]. Resonances at  $\delta$  3.01 ( $d$ ,  $J$  = 12 Hz, 1 H), 4.01 ( $dd$ ,  $J$  = 12, 2 Hz, 1 H) and 3.64 ( $d$ ,  $J$  = 2 Hz, 1 H) in the  $^1\text{H}$  NMR could be assigned to methine protons on C5, C6, and C7 of the B ring of the limonoid skeleton. Acetylation of dysoxylin ( $\text{Ac}_2\text{O}/\text{DMAP}/\text{triethylamine}$  [5]) gave a good yield of the monoacetate (4). The  $^1\text{H}$  NMR of 4 showed that H-7 had undergone an acetylation shift from  $\delta$  3.64 in 1 to 5.0 in 4, and that H-15 had shifted upfield from  $\delta$  3.95 in 1 to 3.69 in 4. These data were consistent with the presence of an alpha hydroxyl substituent at C7 in dysoxylin.

Only four tertiary methyl resonances were apparent in the  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra of 1 (Tables 1 and 2), in contrast to the five expected for the limonoid skeleton and observed in obacunol (2) and obacunone (3). A pair of geminal proton resonances at  $\delta$  4.33 ( $d$ ,  $J$  = 9 Hz) and 3.82 ( $d$ ,  $J$  = 9 Hz) indicated that the fifth methyl residue was connected to C6 via an ether linkage identical to that found in salannin [6] and diacetylvilasinin (5) [7].

The stereochemistry of dysoxylin (1) was determined by an analysis of spin-spin coupling constants and a series of NOE experiments. A 12 Hz vicinal coupling constant between H5 and H6 suggested that ring B was in a chair conformation and that both protons were axial, while a 2 Hz coupling between H6 and H7 implied that H7 was equatorial. Irradiation of the C19 methyl resonance in the monoacetate 4 induced NOE's into Me-30, Me-29 and H6. Irradiation of the C29 methyl resonance induced NOE's into Me-19, H-6, and H-28,  $\beta$ , while irradiation of the C18 methyl resonance induced an NOE into H9. These results are consistent with the stereochemistry shown for 1.

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Table 1.  $^1\text{H}$  NMR data for dysoxylin (1), obacunol (2), dysoxylin acetate (4) and 1,3-diacetylvilasinin (5) (The spectra of 1 and 4 were run in  $\text{CDCl}_3$  at 400 MHz, chemical shifts are reported in ppm from internal TMS)

Compound	1	2*	4	5†
Protons on carbon no.				
1	6.33 d 12 Hz	6.57 d 12 Hz	6.31 d 12 Hz	
2	5.85 d 12 Hz	5.89 d 12 Hz	5.92 d 12 Hz	
5	3.01 d 12 Hz		2.82 d 12 Hz	2.67 d 12.5 Hz
6	4.01 dd 12 and 2 Hz		4.07 dd 12 and 2 Hz	4.14 dd 12.5 and 3 Hz
7	3.64 t 2 Hz	3.52 t 2 Hz	5.01 d 2 Hz	4.18 d 3 Hz
9	2.48 t 8 Hz		2.47 t 8 Hz	2.56 m
15	3.95 s	3.90 s	3.69 s	
17	5.58 s	5.64 s	5.58 s	
18	1.21 s	1.25 s‡	1.20 s	0.84 s
19	1.28 s	1.30 s‡	1.29 s	0.97 s
21	7.39 m	7.49 d 1 Hz	7.41 m	7.36 m
22	6.33 d 1 Hz	6.42 t 1 Hz	6.34 d 1 Hz	6.27 m
23	7.39 m	7.49 d 1 Hz	7.41 m	7.24 m
28	3.82 d 9 Hz	1.25 s	3.77 9 Hz	3.59 d 7 Hz
	4.33 d 9 Hz		4.24 9 Hz	3.56 d 7 Hz
29	1.60 s	1.43 s‡	1.55 s	1.18 s
30	1.08 s	1.05 s‡	1.17 s	1.11 s

\* Data from ref. [3].

† See ref. [7] for complete data.

‡ Unassigned.

Dysoxylin (1) was hydrolysed with 0.05 M NaOH in dioxane to give the dihydroxy acid 6, which was methylated with diazomethane to give the ester 7. Acetylation of ester 7 ( $\text{Ac}_2\text{O}$ /pyridine) gave a mixture of mono- and diacetates 8 and 9. The spectral data observed for all the transformation products were consistent with the proposed structures.

## EXPERIMENTAL

**Isolation and extraction.** Fresh leaves of *Dysoxylum richii* were collected from Suva, Fiji in December 1985. A voucher specimen of this plant is deposited at the South Pacific Regional Herbarium, Institute of Natural Resources, University of the South Pacific, Suva, Fiji. The air-dried leaves (300 g) were soaked in MeOH (1.5 l) for 5 days. The MeOH extract was concentrated to a gum,  $\text{H}_2\text{O}$  was added and the resulting suspension was extracted successively with petrol (bp 60–80°) and  $\text{CHCl}_3$ . The concentrated  $\text{CHCl}_3$  extract was dissolved in  $\text{EtOAc}$  (10 ml), soaked over alumina (10 g), filtered, and concentrated to give a gum (2.4 g). Fractionation of the gum by flash chromatography on silica gel using mixtures of cyclohexane– $\text{EtOAc}$  gave crude dysoxylin (1) (0.37 g). Preparative TLC (1:1 toluene/ $\text{EtOAc}$ ) of the crude material gave 1 as solid, mp 247–251°,  $[\alpha]_D^{25} + 72.5^\circ$  ( $\text{CHCl}_3$ ; c 0.15);  $\text{C}_{26}\text{H}_{30}\text{O}_8$  EIMS  $[\text{M}]^+$  470.1937 (dev. –0.4 mmu);  $^1\text{H}$  NMR see Table 1;  $^{13}\text{C}$  NMR see Table 2.

**Dysoxylin acetate (4).** To a stirred mixture of 1 (20 mg) and  $\text{NEt}_3$  (0.5 ml) was added 4-(*N,N*-dimethylamino)pyridine (DMAP) (3 mg). The reaction mixture was heated to 60° for 3 hr.

After cooling, the reaction mixture was diluted with water (5 ml), then extracted with  $\text{CHCl}_3$ . The organic layer was washed sequentially with aq. HCl (5%) and aq.  $\text{NaHCO}_3$  (5%) before drying and concentration *in vacuo* to give dysoxylin acetate (4) as a solid, mp 235–245°;  $\text{C}_{28}\text{H}_{32}\text{O}_9$  EIMS  $[\text{M}]^+$  512.2050 (dev. +0.3 mmu);  $^1\text{H}$  NMR see Table 1;  $^{13}\text{C}$  NMR see Table 2.

**Dihydroxy acid.** To a solution of 1 (3 mg) in dioxane (0.5 ml) was added 0.05 M NaOH (0.5 ml). The solution was refluxed for 30 min, then warmed at 50° for 90 min. Work-up gave the dihydroxyacid 6.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ) 1.05 (3H, s, Me-30), 1.27 (3H, s, Me-18), 1.34 (3H, s, Me-19), 1.45 (3H, s, Me-29).

**Methyl ester.** Dihydroxyacid 6 was methylated with diazomethane to give the methyl ester 7.  $\text{C}_{27}\text{H}_{34}\text{O}_9$  EIMS  $[\text{M}]^+$  502.2110 (dev. 0.7 mmu);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ) 1.07 (3H, s, Me-30), 1.22 (3H, s, Me-18), 1.35 (3H, s, Me-19), 1.45 (3H, s, Me-29), 3.75 (3H, s, COOMe).

**Monoacetate and diacetate.** Acetylation of 7 using  $\text{Ac}_2\text{O}$ –pyr. (room temp. for 24 hr) gave the monoacetate 8 and the diacetate 9 as gums. 8:  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ) 1.11, 1.21, 1.32 and 1.45 (3H each, s, Me-30, Me-18, Me-19 and Me-29) 2.20 (3H, s, COMe), 3.73 (3H, s, COOMe), 5.00 ppm (1H, d,  $J$  = 2 Hz, H-7); 9:  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ) 1.12, 1.23, 1.33 and 1.5 (+ $\text{H}_2\text{O}$ ) (3H each, s, Me-30, Me-18, Me-19 and Me-29), 1.93 and 2.20 (3H each, s, COMe), 3.70 ppm (3H, s, COOMe).

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Table 2.  $^{13}\text{C}$  NMR data. All spectra were run in  $\text{CDCl}_3$  (Chemical shifts are in ppm from internal TMS)

Carbon	1*	3†	4	5‡
1	149.8	156.8	149.3	72.28
2	119.4	122.7	119.9	27.66
3	167.7§	166.7	166.8§	71.76
4	84.9	84.0	84.4	42.32
5	52.4	53.3¶	54.3	39.62
6	77.5	39.0	76.0	72.90
7	69.6	207.5	71.0	74.01
8	43.3	52.9	43.3	45.80
9	40.4	49.1	42.0	33.62
10	44.8	43.1	44.5	39.20
11	15.4	17.0	15.4	15.21
12	26.0	32.6	26.0	33.02
13	38.5	37.3	38.8	47.36
14	69.6	65.1	69.5	159.88
15	58.1	57.2¶	56.1	120.75
16	165.6§	166.9	165.5§	34.35
17	78.1	77.9	78.0	51.58
18	17.5	16.4	17.7§	21.16
19	17.2	19.4	17.3§	26.20
20	120.5	120.0	120.2	124.52
21	143.0	143.1	143.1	139.76
22	109.9	109.7	109.8	111.06
23	141.2	140.9	141.2	142.62
28	81.3	21.0	81.3	77.88
29	23.5	26.7	23.4§	15.37
30	17.9	32.0	18.5§	19.47
MeCO	—	—	21.0	21.16, 21.09
MeCO	—	—	170.0	169.97, 170.28

\*Assignments were based on comparison to literature values for model compounds, APT and one bond HETCOR.

†Data from ref. [4] (see also [8]).

‡Data from ref. [7].

§Values in same vertical column may be reversed.

||Unassigned.

¶Assignments are reversed in ref. [4].

#### REFERENCES

1. Taylor, D. A. H. (1984) in *Progress in the Chemistry of Organic Natural Products* (Herz, W., Grisebach, H. and Kirby, G. W., eds) Vol. 45, p. 1. Springer, New York.
2. Singh, S., Garg, H. S. and Khanna, N. M. (1976) *Phytochemistry* 15, 2001.
3. Dreyer, D. L. (1968) *J. Org. Chem.* 33, 3577.
4. Dreyer, D. L. and Bennett, R. D. (1976) *Tetrahedron* 32, 2367.
5. Hasner, A., Krepški, L. R. and Alexanian, V. (1978) *Tetrahedron* 34, 2069.
6. Henderson, R., McCrindle, R., Melera, A. and Overton, K. H. (1968) *Tetrahedron* 24, 1525.
7. Kraus, W. and Cramer, R. (1981) *Liebigs. Ann. Chem.* 181.
8. Kubo, I., Tanis, S. P., Lee, Y-W., Miura, I., Nakanishi, K. and Chaypa, A. (1976) *Heterocycles* 5, 485.